## Cattle and Sheep Skin Permeability: A Comparison of Frozen and Reconstituted Skin with that of Fresh Skin

**Keyphrases**  $\square$  Permeability—cattle and sheep skin, comparison of frozen and reconstituted skin with fresh skin  $\square$  Skin—permeability in cattle and sheep, comparison of frozen and reconstituted skin with fresh skin  $\square$  Topical drug delivery—cattle and sheep skin permeability, comparison of frozen and reconstituted skin with fresh skin

## To the Editor:

It is known that freezing and reconstituting (*i.e.*, thawing) excised human skin does not significantly change the barrier properties it presents to the passive diffusion of drug molecules (1). This knowledge has led to substantial advances in the development of topical drug delivery systems for humans, because potentially useful systems can be identified from the results of *in vitro* screens using frozen and reconstituted human skin (2).

This communication presents data to establish that freezing and reconstituting cattle and sheep skin does not significantly alter their permeability.

It has previously been established (3) that the outer 1 mm of frozen and reconstituted cattle skin (*i.e.*, the stratum corneum, the viable epidermis, the papillary region of the dermis, and a portion of the reticular region of the dermis) acts as an homogeneous barrier to diffusing levamisole molecules. A corollary to this finding is that, for skin samples with thicknesses up to 1 mm, the product of the permeability constant ( $k_p$  in centimeters per minute) and the skin thickness (r in centimeters) (*i.e.*,  $k_pr$ ) is a constant. A similar relationship has been observed for penetration of levamisole through the skins of Merino sheep (4).

To establish the relative permeabilities of fresh and reconstituted cattle and sheep skins, the following experiments were conducted.

Skin was harvested from an 8–9-month Shorthorn-Hereford cross calf in early spring and from a 12–18-month Merino cross ewe in mid-winter. The sheep was shorn and then finely clipped<sup>1</sup> and the calf finely clipped<sup>1</sup> immediately before skin samples were removed with a dermatome<sup>2</sup> set at 1.1 mm.

The permeability of fresh skin and skin that had been stored at  $-30^{\circ}$  for 5–7 days and then thawed to levamisole from a 0.85% solution in an aqueous pH 8.9 buffer was determined using methods identical to those described previously (3).

Values of  $k_p r$  for levamisole penetrating through fresh and reconstituted skins are given in Table I.

There was no significant difference (at the 1 or 5% levels)

Table I—Permeability of	Cattle and Sheep	Skins to Levamisole <sup>a</sup>
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Animal	Skin Thickness, cm	$10^{6} k_{p} r,$ cm <sup>2</sup> min <sup>-1</sup>
Sheep	Fresh 0.055 0.066 0.070	6.0 7.7 10.7
	0.071 Frozen	9.2
Sheep	0.077 0.077 0.077 Fresh	7.2 6.5 8.4
Calf	0.069 0.071 0.076 0.092 0.093	27.7 26.8 30.9 33.7 31.7
Calf	Frozen 0.065 0.069 0.071 0.082 0.086 0.091	26.5 25.6 29.3 29.5 30.2 33.4

 $^a$  From a 0.85% solution in an aqueous pH 8.9 buffer and water-bath temperature of 37°.

between the values of  $k_p r$  for fresh and reconstituted cattle or sheep skin.

Although no relationship has been established between in vitro and in vivo permeability for cattle and sheep skins, it seems unlikely that live skin, with its blood supply and sweat and sebum secretions, would be appreciably less permeable than excised skin, unless the latter was damaged by the receptor solution.

Consequently, it is concluded that screens for cattle and sheep skin permeability that employ reconstituted skin (5-7) and utilize normal saline as a receptor phase have the potential to provide useful information to developers of veterinary topical dosage forms.

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<sup>&</sup>lt;sup>1</sup> Andis R400 Oster A5 Clippers.

<sup>&</sup>lt;sup>2</sup> Brown Electro Dermatome, model 902.